

Using GeneLab for VESGEN Systems Analysis of Vascular Phenotypes from Stress and Other Signaling Pathways

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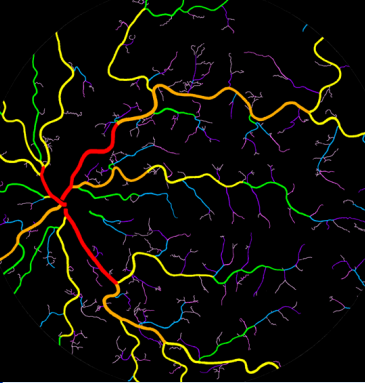
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One fundamental requirement shared by humans with all higher terrestrial life forms, including other vertebrates, insects, and higher land plants, is a complex, fractally branching vascular system. NASA's VESSEL GENERATION Analysis (VESGEN) software maps and quantifies vascular trees, networks, and tree-network composites according to weighted physiological rules such as vessel connectivity, tapering and bifurcational branching. According to fluid dynamics, successful vascular transport requires a complex distributed system of highly regulated laminar flow. Microvascular branching rules within vertebrates, dicot leaves and the other organisms therefore display many similarities. A unifying perspective is that vascular patterning offers a useful readout of molecular signaling that necessarily integrates these complex pathways. VESGEN has elucidated changes in vascular pattern resulting from inflammatory, developmental and other signaling within numerous tissues and major model organisms studied for Space Biology.

For a new VESGEN systems approach, we analyzed differential gene expression in leaves of *Arabidopsis thaliana* reported by GeneLab (GLDS-7) for spaceflight. Vascular-related changes in leaf gene expression were identified that can potentially be phenocopied by mutants in ground-based experiments. To link transcriptional, protein and other molecular change with phenotype, alterations in the spatial and dynamic dimensions of vascular patterns for *Arabidopsis* leaves and other model species are being co-localized with signaling patterns of single molecular expression analyzed as information dimensions. Previously, *Drosophila* microarray data returned from space suggested significant changes in genes related to wing venation development that include EGF, Notch, Hedgehog, Wingless and Dpp signaling. Phenotypes of increasingly abnormal ectopic wing venation in the (non-spaceflight) *Drosophila* wing generated by overexpression of a Notch antagonist were analyzed by VESGEN. Other VESGEN research applications include the mouse retina, GI and coronary vessels, avian placental analogs and translational studies in the astronaut retina related to health challenges for long-duration missions.

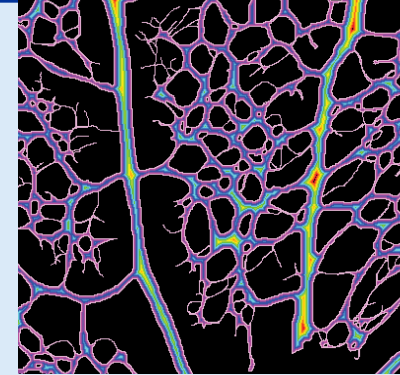
Research partially supported by Ames Center Innovation Award to PP-W.



Human Retina

NASA'S VESGEN Software

Research Discovery Tool
for Fundamental and Translational
Space Biology Research



Mouse Retina

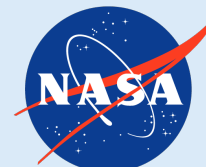
Using NASA's GeneLab for VESGEN Systems Analysis of Vascular Phenotypes from Stress and Other Signaling Pathways

Patricia Parsons-Wingerter
Space Biosciences Research Branch

VESGEN Patents Pending

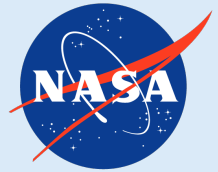


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Collaborators

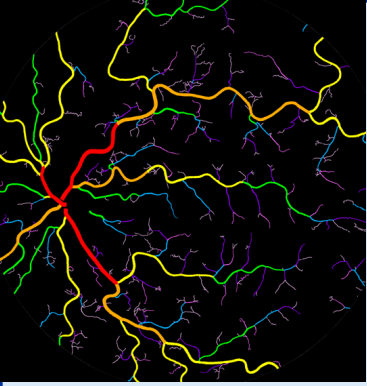
Alexander Weitzel, USRA Summer 2016 Internship, NASA Ames Research Center & Grand Valley State University,

Ruchi Vyas and Matthew Murray, MORI Associates and Blue Marble Space Institute, NASA Ames Research Center

Mary B. Vickerman, Software Systems, NASA Glenn Research Center

Sharmila Bhattacharya Space Biosciences Research Branch, NASA Ames Research Center

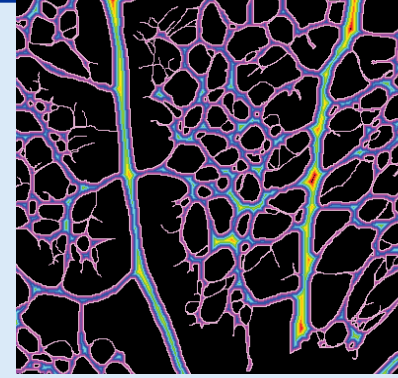
Sarah E. Wyatt, Environmental and Plant Biology, Ohio University, Athens OH



Human Retina

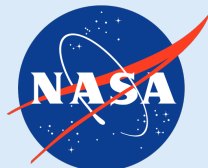
NASA'S VESsel GENeration Analysis [VESGEN] Software

Mapping and Quantification of Branching Vascular Patterns from Physiological Branching Rules



Mouse Retina

- Vascular trees, networks, and tree-network composites from set of weighted parameters for vessel connectivity, tapering and bifurcational branching
- Requirements of fluid dynamics for laminar flow
Aqueous vascular transport by complex distributed system of fractal-based bifurcational branching
- Microvascular rules for fractal-based branching within humans, vertebrates, insects and dicot leaves therefore display many similarities



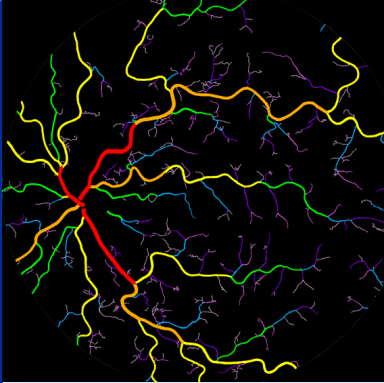


Parsons



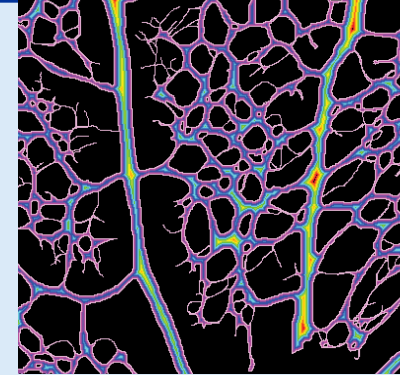
NASA





Human Retina

VESGEN Analysis for Fundamental Space Biology Research with Translational Applications to Astronaut Health and Countermeasures



Mouse Retina

Vascular Trees

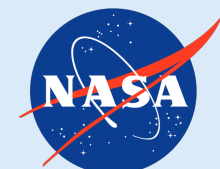
Retinas of Astronauts and Human Bed Rest; Diabetic Retinopathy
Mouse/Avian Coronary Vessels, Chorioallantoic Membrane (CAM), Yolk sac

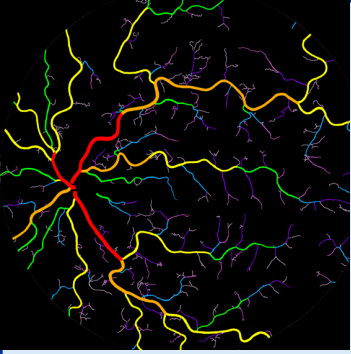
Vascular Networks

Mouse Intestinal Inflammation, CAM Lymphatic Vessels, Abnormal Mouse Corneal Angiogenesis, *Drosophila* (Fruitfly) Wing

Vascular Tree-Network Composites

Mouse Postnatal Retina
Early Embryonic Coronary Vessels, *Arabidopsis* Leaf Venation

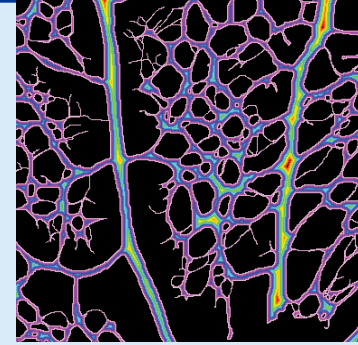




Human Retina

VESGEN

Analysis of Vascular Patterns



Mouse Retina

Research Hypothesis

Vascular patterning offers useful readout of molecular signaling that necessarily integrates crosstalk among complex signaling pathways

Fractal-Based Physiological Branching Rules

from fluid mechanics, anatomy, microscopic observations

Mapping and Quantification by Multiparametric Weighted Analysis

Fractal Dimension, D_f

Vessel Number Density, N_v

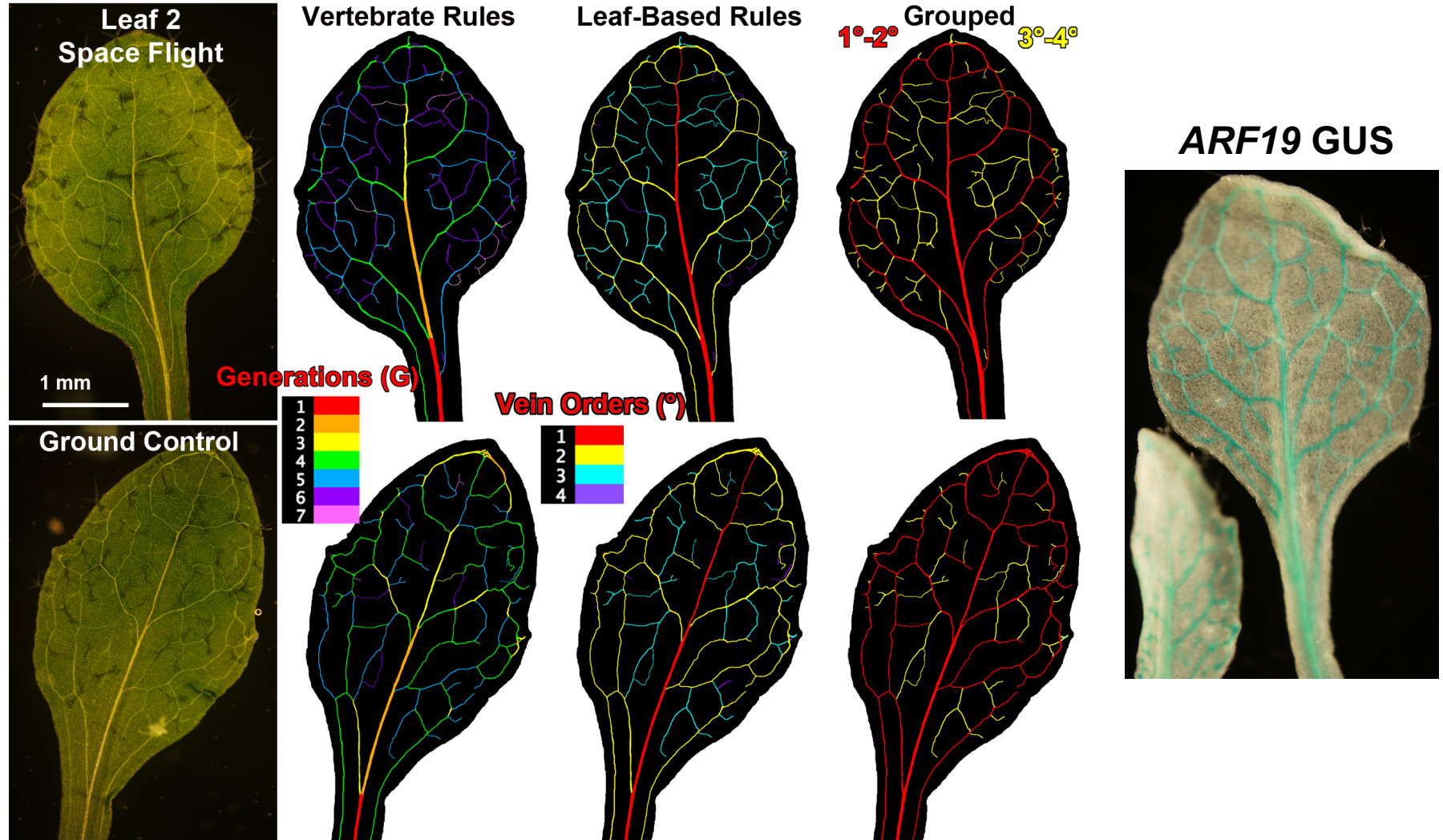
Vessel Length Density, L_v

Vessel Diameter, D_v

Branchpoint + Endpoint Densities, $Br_v + E_v$

Arabidopsis leaves from ISS: STS-130

Juvenile Leaf 2



VESGEN mapping of Arabidopsis leaf venation with bioinformatic analysis

Kang & Dengler, *Int J Plant Sci*
 165(2): 231 **2004** ©University of Chicago Press

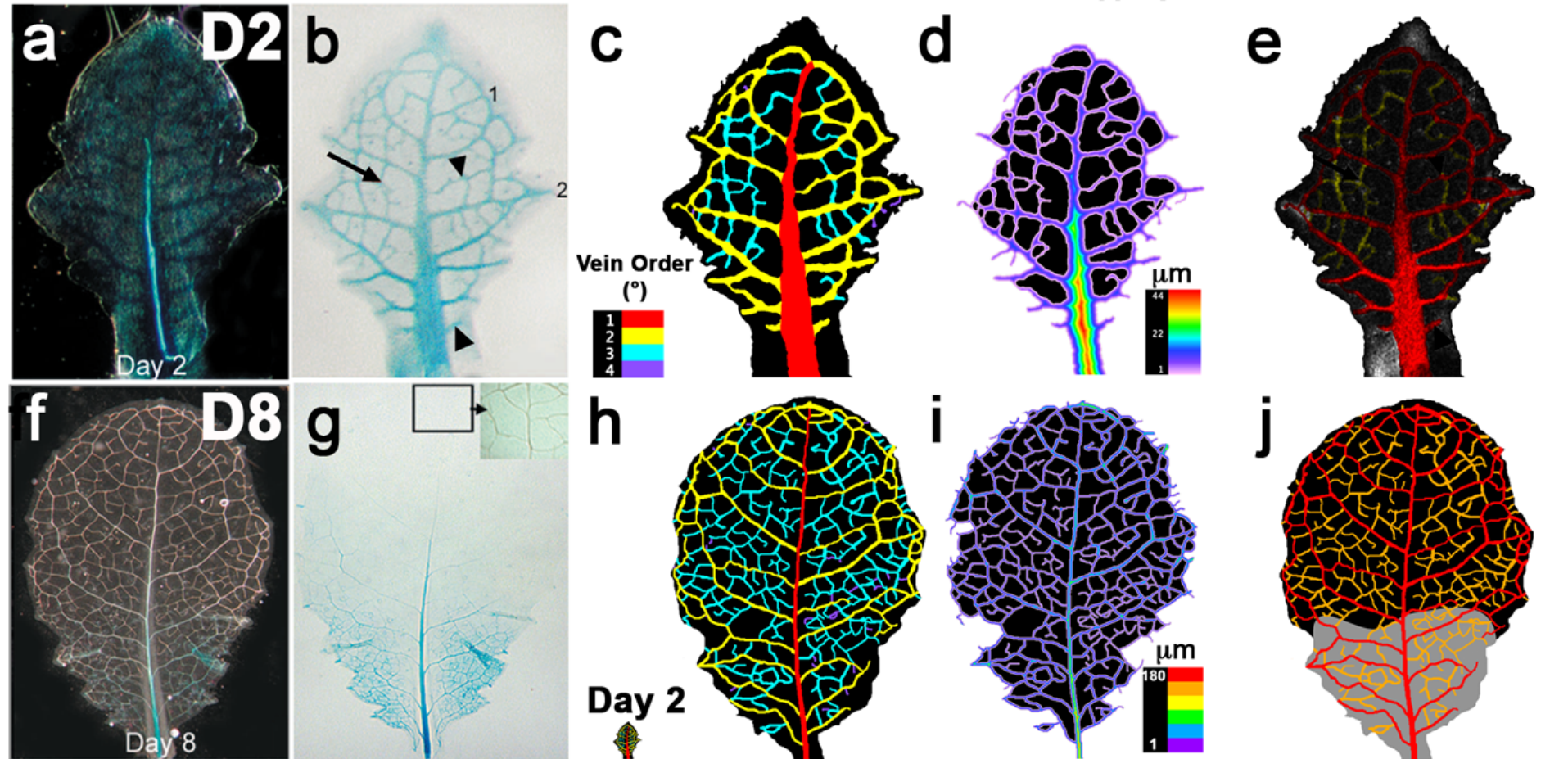
**Differentiated
Xylem**

***AtHB8::GUS*
Expression**

Taxonomy & Phylogeny:
**Grouping by Venous
Branching Orders**

**Venous Diameter by
Distance Mapping**

Integrative Bioinformatics:
***AtHB8::GUS* by Structural &
Reticulate Vein Grouping**





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The Arabidopsis spaceflight transcriptome: a comparison of whole plants to discrete root, hypocotyl and shoot responses to the orbital environment



2 datasets available for download here:

ISA-TAB Metadata file
Raw Data File

GeneLab Accession Number	GLDS-7			
Source Accession Number	E-MTAB-1264			
Contacts	Name	Role	Organization	Email
	Robert Ferl	Investigator	University of Florida	robferl@ufl.edu
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	Agata Zupanska	Submitter	University of Florida	zupanska@ufl.edu

Space Grown *Arabidopsis* with Microarray Data from GeneLab: Identification of Genes Important in Vascular Patterning

(A Weitzel, P Parsons, S Wyatt; ASGSR 2016)

- Analysis of transcriptomic data from space flight and ground control leaves identified differential expression of 22 genes, of which seven may be related to plant vasculature
- Two gene clusters suggest there may be phenotypic changes in leaf venation resulting from development in microgravity
 - KISS ME DEADLY [KMD]** coding F-box genes
 - NAM**, **ATAF1/2**, and **CUC2 [NAC]** related genes
- Vascular-related changes in leaf gene expression can potentially be phenocopied by mutants in ground-based experiments and corroborated by VESGEN analysis
- Genetic, transcriptional and other molecular changes reported by GeneLab can be mapped to vascular phenotypes by VESGEN by bioinformatic co-localization of single molecular expression

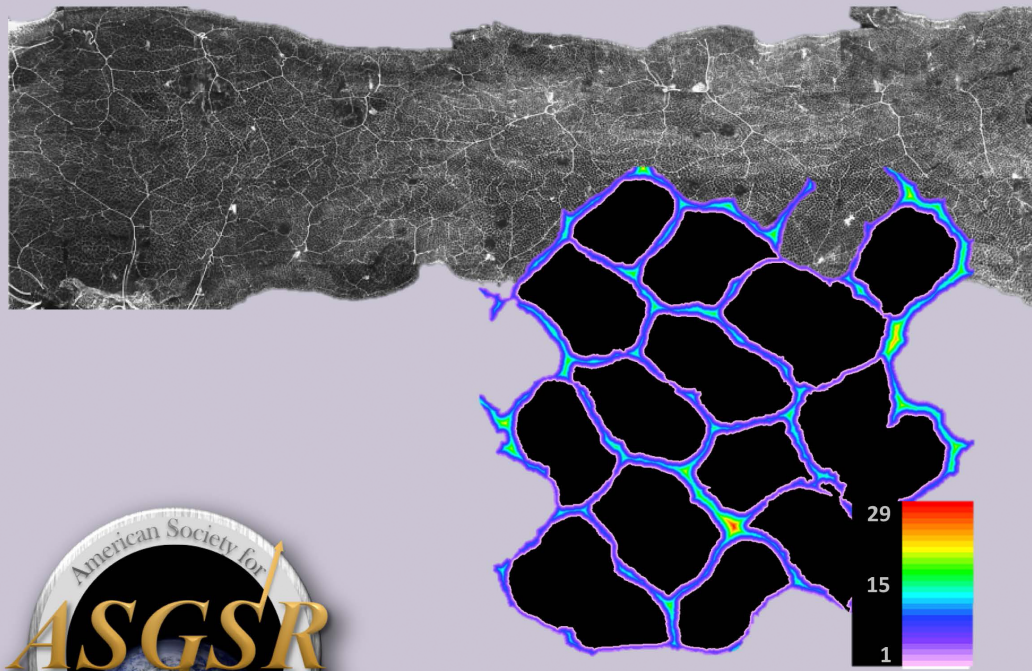


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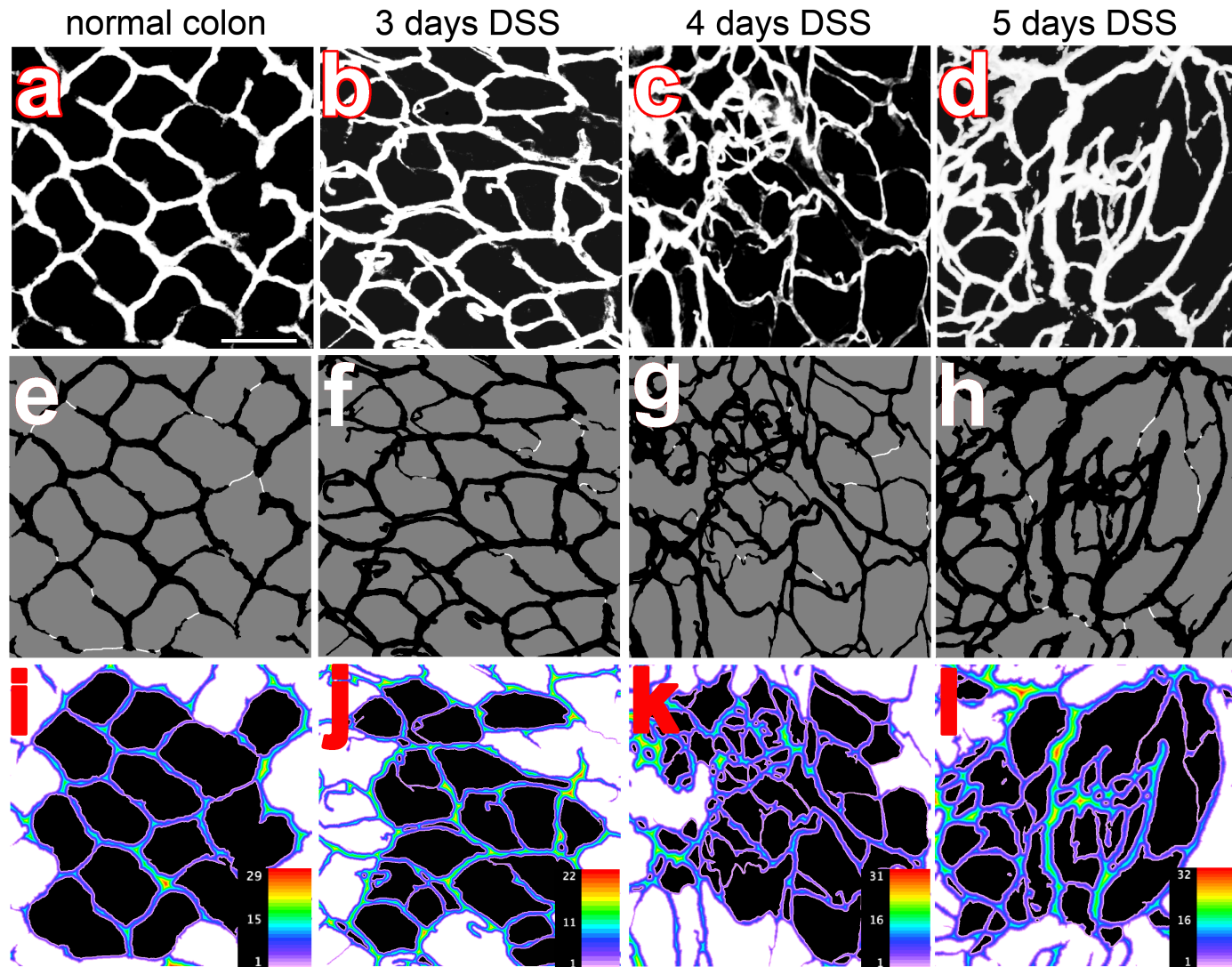


Gravitational and Space Biology

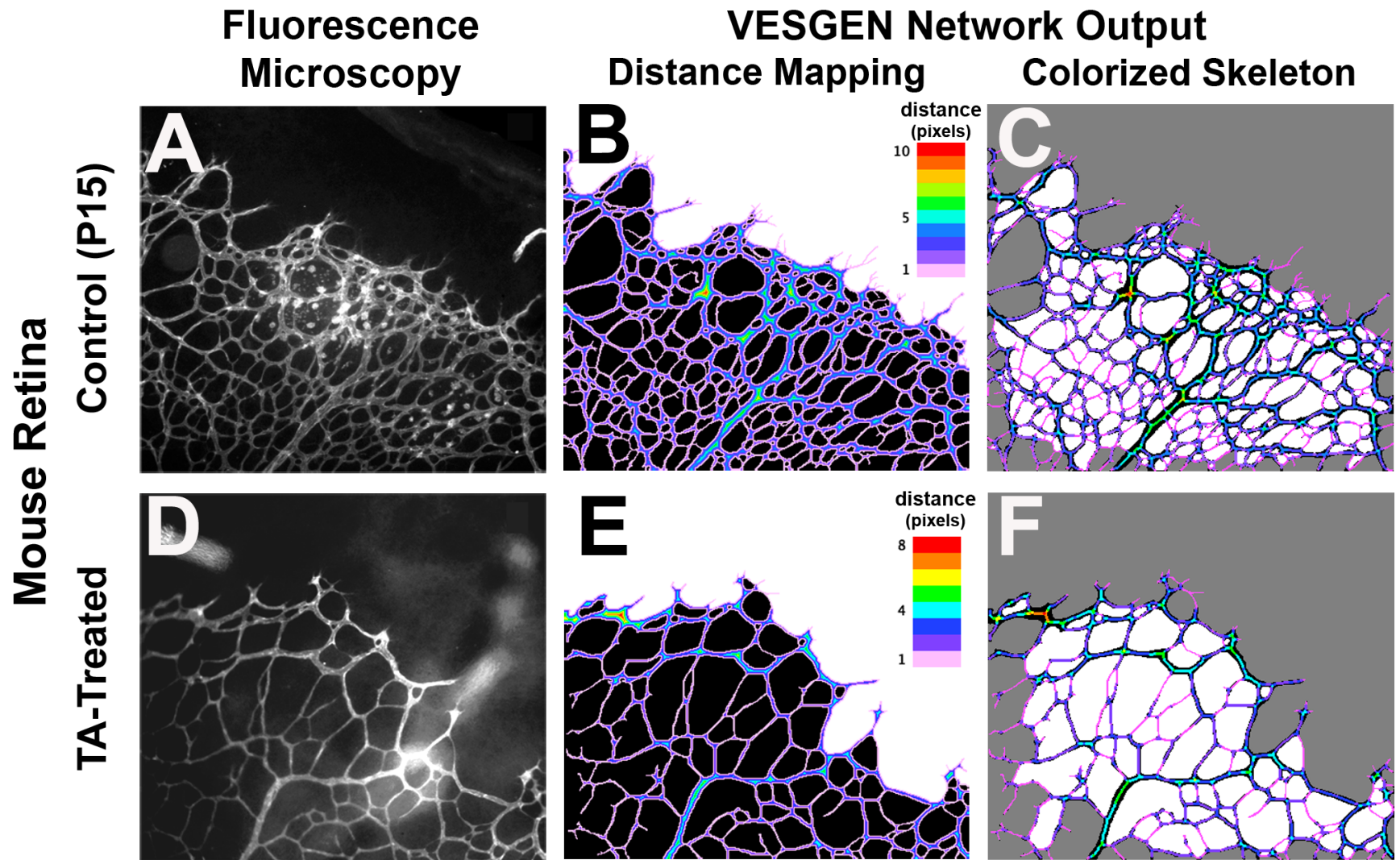
Publication of the American Society for Gravitational and Space Research



VESGEN mapping of vascular networks for progressive GI inflammation progression with mouse model

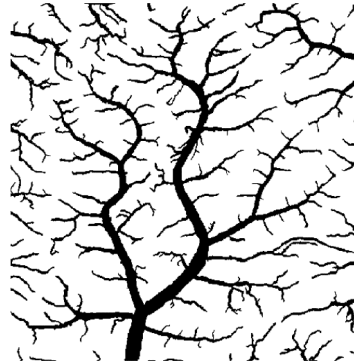
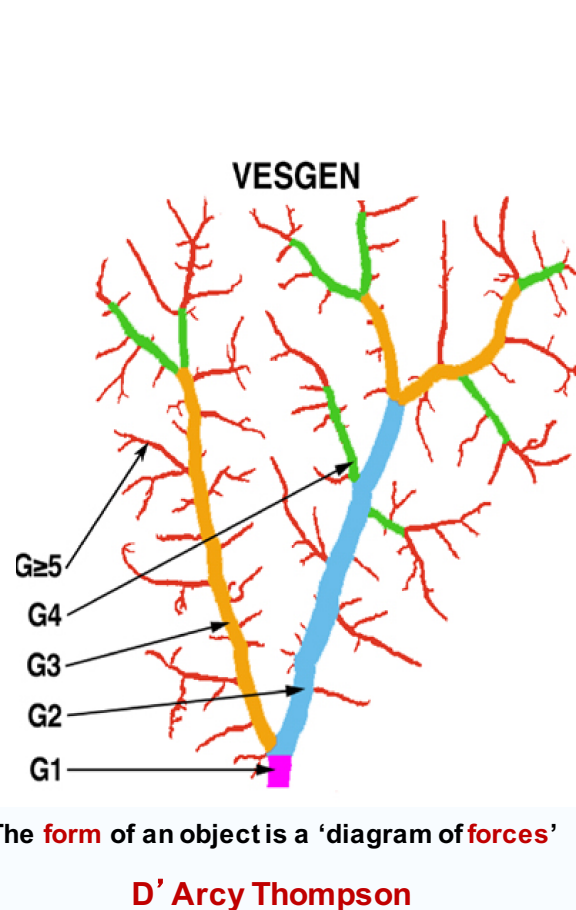


Vascular Networks in Transgenic Mouse Retina

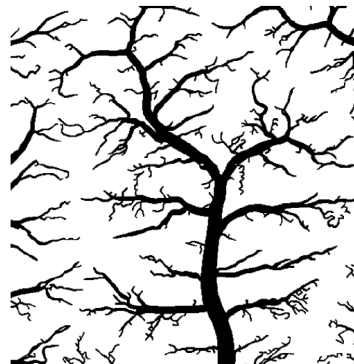
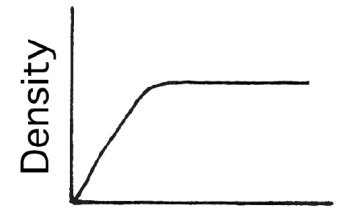


with J Sears & Q Ebrahim (Cole Eye Institute), from Vickerman et al, *Anat Rec* 292(3), 2009

VESGEN Hypothesis ‘Fingerprint’ or ‘Signature’ Vascular Pattern as Useful Integrative Readout of Complex Molecular Signaling Pathways



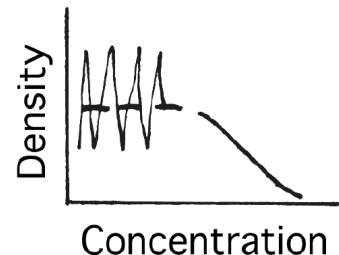
bFGF as Simple Stimulator
Arterio Thromb Vasc Biol 20 (2000)



VEGF as Vascular Complexity Factor: Phenotypic Readout with eNOS Signaling
Microvascular Research 72 (2006)



TGF- β 1 as Simple Inhibitor but Complex Potentiator
Microvascular Research 59 (2000)

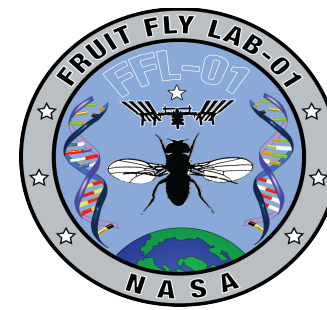
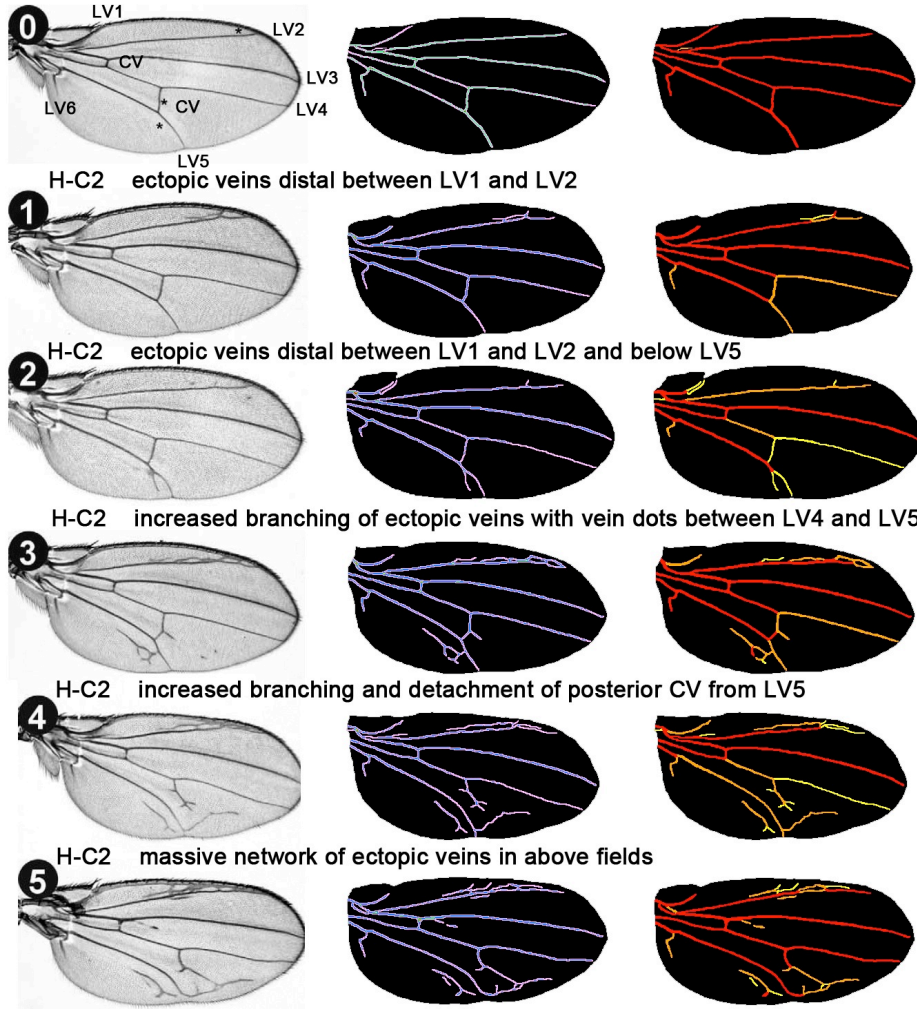


Fruitfly (*Drosophila melanogaster*): Major Genetic Model Organism

Mapping by VESGEN: Hairless (H-C2) overexpression induces phenotypic series of increasing ectopic wing venation

6 Longitudinal Veins (LV) with anterior and posterior Cross Veins (CV)

Wildtype, no heat shock * where veinlets may arise after heat shock in wt or H-C2



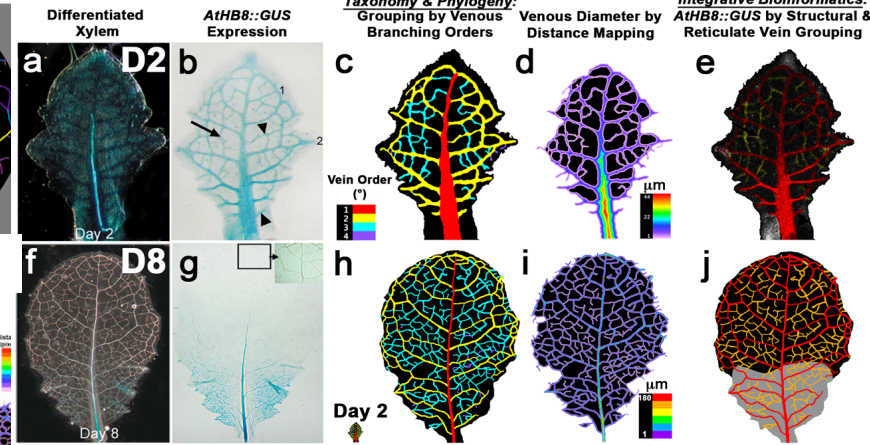
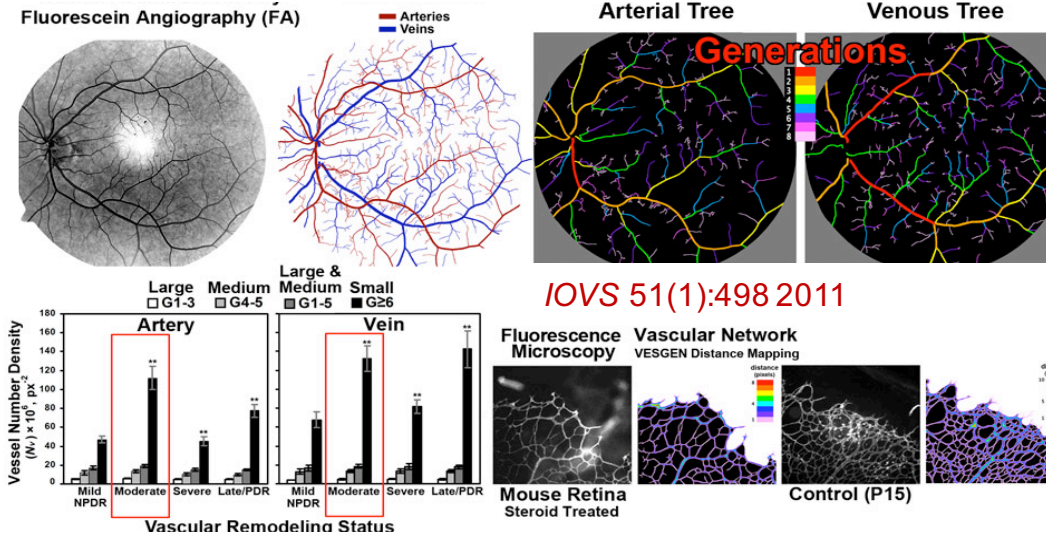
Expression of Genes Involved in *Drosophila* Wing Morphogenesis and Vein Patterning Are Altered by Spaceflight

Parsons, Hosamani, Vickerman, Bhattacharya
Grav Space Res 3(2):54-64 2015, ASGSR 2012

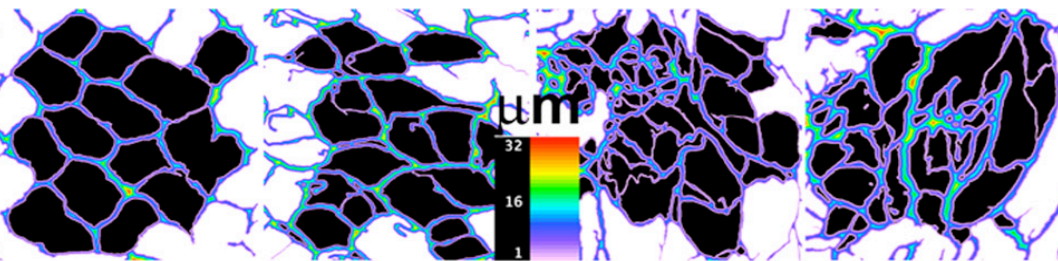
VESGEN Analysis of Vascular Patterning for Fundamental Space Biology with Translational Applications to Astronaut and Terrestrial Health

Insights into progression of visually impairing disease

GSR 2(1):68 2014

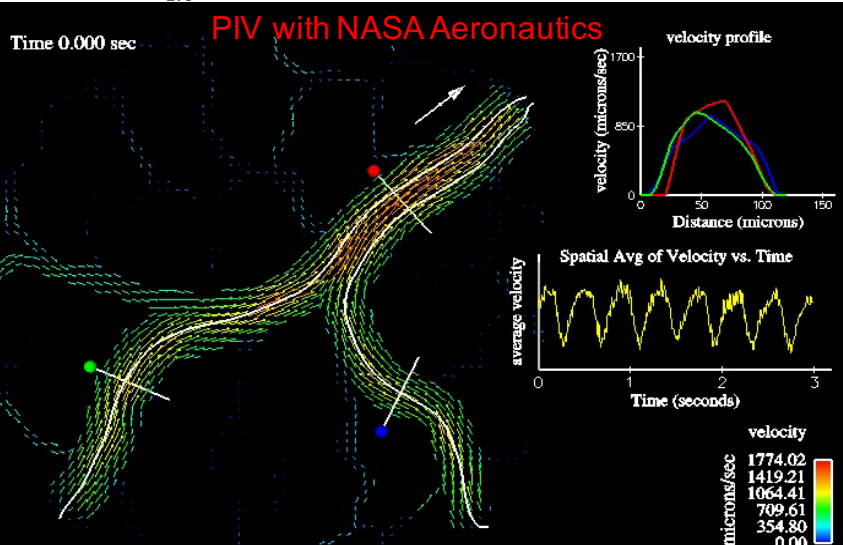
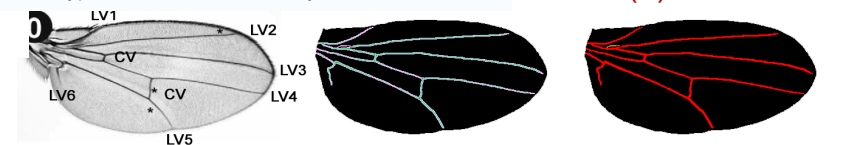


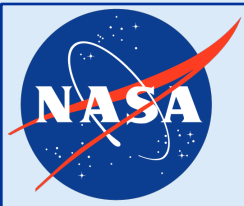
GSB 26(2):2 2012, PloS One 8(5):e64227 2013



Wildtype *where veinlets may arise in H-C2 mutant

GSR 3(2):54-64 2015





Acknowledgments



Glenn Research Center Patricia Keith, Mark Wernet, numerous student interns

Veteran's Administration & Ohio Aerospace Institute

Krishnan Radhakrishnan

University of Florida Gainesville, Horticultural Center

Anna-Lisa Paul, Rob Ferl

Massachusetts General Hospital, Harvard Medical School

Hans-Christian Reinecker & colleagues

Cleveland Clinic Foundation

Cole Eye Institute— Peter Kaiser, Jonathan Sears, Quteba Ebrahim

Lerner Research Institute— Paul DiCorleto, Unni Chandrasekharan

Case Western Reserve University

Michiko Watanabe, Monica Montano, Karunamuni G

Supported by NASA Ames and Glenn Center Innovation Fund Awards,
NASA Space Life and Physical Sciences Program, National Institutes of Health R01 EY017529

